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An increase in the energy coupling capacity of submitochondrial particles in the presence of lanthanides

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Abstract

NADH and succinate oxidase activities of inside-out submitochondrial particles treated with excess oligomycin are inhibited by lanthanides (La³⁺ and Dy³⁺). Inhibition by both oligomycin and oligomycin plus lanthanides is completely relieved by an uncoupler. The respiratory control, measured as the stimulation of NADH or succinate oxidation caused by the addition of uncoupler to the oligomycin-treated particles, is thus increased in the presence of lanthanides. The coupling effect of lanthanides is completely prevented and rapidly reversed by excess of EDTA. La³⁺ increases the extent of the aerobic energy-linked succinate-supported NAD⁺ reduction catalyzed by the oligomycin-treated submitochondrial particles. Lanthanides seem to be a useful tool to increase the energy coupling capacity of the submitochondrial particles.

Key words: Lanthanide; Energy coupling; Submitochondrial particle

1. Introduction

Respiratory control, as measured by stimulation of respiration by an uncoupler, has been demonstrated in certain submitochondrial particles (SMP) treated with F₀-directed energy transfer inhibitors, such as oligomycin [1,2], DCCD [3], chloromercurinitrophenol [4], or with a number of 'coupling factors' derived from the resolved mitochondrial preparations [5,6]. The ability of oligomycin and DCCD to induce respiratory control provides a simplified system for the analysis of the kinetics and thermodynamics of the respiratory chain components under the energized state, i.e. when the free energy of the intramolecular redox reactions within the individual complexes is in apparent equilibrium with $\Delta \overline{\mu}_{H^+}$. It is anticipated that some of the redox components directly involved in the vectorial H⁺ translocation may only be detected under such an energized state. For example, rapidly relaxing Complex I-associated ubisemiquinone(s) is only seen in tightly coupled SMP [7].

One general problem in studies with resolved SMP is that even in the presence of oligomycin or DCCD the preparations are never absolutely coupled. The reasons for this are not clear: the heterogeneity of inside-out SMP, proton leakage through the phospholipid bilayer, unspecific protein-mediated proton conductivity, and possible mistakes in the operation of the proton pumps may contribute to the respiration in the presence of the

Abbreviations: SMP, submitochondrial particles; BSA, bovine serum albumin; DCCD, N,N'-dicyclohexylcarbodiimide; F_0 , membrane sector of the mitochondrial F_0 - F_1 ATPsynthase; Complex I, NADH-ubiquinone oxidoreductase (EC 1.6.99.3); CCCP, carbonylcyanide-m-chlorophenylhydrazone

energy-transfer inhibitors. A search for the factors which would increase the energy coupling capacity of SMP thus seems to be worthwhile. In this report we will show the lanthanides (La⁺³ and Dy⁺³) are capable of increasing the respiratory control and stimulation of the energy-linked reverse electron transfer in the oligomycin-treated SMP.

2. Materials and methods

Bovine heart submitochondrial particles were prepared as described [8] and stored in liquid nitrogen. NADH oxidase activity was measured spectrophotometrically as a decrease in absorption at 340 nm ($\varepsilon_{\text{340, mM}} = 6.22$). Succinate oxidase was measured with oxygen electrode. Protein content was determined by the biuret procedure [9]. BSA (Fraction V), oligomycin and CCCP were from Sigma (USA), LaCl₃·6H₂O and DyCl₃·4H₂O were from Alfa Products (USA), NADH was from Reanal (Hungary), Tris was from Serva (Germany). Other chemicals were of the purest grade commercially available.

3. Results

In agreement with classical observations [1,2] oligomycin at a concentration inhibiting more than 90% of ATP-ase activity of SMP causes an inhibition of respiration with NADH as the substrate, and this inhibition is relieved by an uncoupler (Fig. 1, trace A). Respiration in the presence of oligomycin was further decreased by LaCl₃ and uncoupler caused the activation of NADH oxidase up to the level observed without lanthanide (Fig. 1, trace B). The net result caused by lanthanide was an increase in the oligomycin-induced respiratory control. Table 1 summarizes data illustrating this phenomenon. In different batches of SMP routinely prepared in this laboratory [8] the respiratory control ratios, as measured by stimulation of respiration by an uncoupler after treatment with oligomycin, are 3:8 with NADH and 3:5 with

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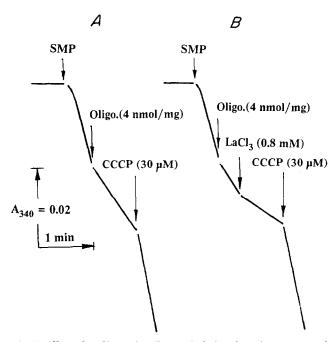


Fig. 1. Effect of LaCl₃ on the oligomycin-induced respiratory control. The assay cuvette (2 ml) contained 0.2 M sucrose, 50 mM Tris-acetate (pH 7.5), BSA (1 mg/ml) and 0.1 mM NADH. The reaction was initiated by the addition of submitochondrial particles (12 μ g/ml). Further additions are indicated.

succinate. Both La⁺³ and Dy⁺³ increased respiratory control in loosely coupled SMP with either NADH (Exps. 1 and 2) or succinate (Exp. 3). It should be noted that high concentrations of succinate (20 mM) prevented the coupling effect of La⁺³ (Exp. 1, sample 3) presumably due to chelation of the lanthanide cation [10]. The same phenomenon was observed with EDTA, which completely abolished the coupling effect of La⁺³ (Fig. 2) and Dy⁺³ (Table 1; Exp. 2, sample 3). Nevertheless, the coupling effect of La⁺³ was observed with succinate when the concentration of the latter was decreased to 1 mM (Table 1; Exp. 3). The coupling effect of La⁺³ was concentration-dependent (saturation at ~0.9 mM) and completely reversed by the slight molar excess of EDTA (Fig. 2).

The observations reported may be interpreted in terms of the coupling effect of lanthanides on energy conservation in the respiratory chain. Therefore, it might be expected that the energy-linked reactions catalyzed by SMP would be stimulated by lanthanides. In an attempt to relate the inhibitory effect of La⁺³ and Dy⁺³ on respiration to their coupling capacity, the aerobic succinate-supported reverse electron transfer was examined. In agreement with our previous data [8], oligomycin-treated SMP catalyzed NAD⁺ reduction at the expense of energy produced by the succinate oxidation (Fig. 3). With limited NAD⁺ the steady-state mass action ratio of 14 for NAD⁺/NADH couple was established. The addition of La⁺³ caused an additional reduction in NAD⁺ (NAD⁺/NADH ratio equal to 6 was reached) which was com-

pletely reversed by 10 mM malonate or by an uncoupler (not shown). The results obtained unambiguously suggest that lanthanides increase $\Delta \overline{\mu}_{H^+}$ across the coupling membrane without any inhibitory effect on the respiratory chain.

It was shown in separate experiments that neither Ruthenium red (30 nM) nor Ca⁺² (1 mM) and another polyvalent cation, spermine (200 μ M), had any significant effect on respiration and the reverse electron transfer catalyzed by the oligomycin-treated SMP. Lanthanides were unable to decrease respiration of the tightly coupled SMP having the respiratory control ratio of 7:9 in the presence of oligomycin with NADH as a substrate.

4. Discussion

It is obvious that an artificial improvement of energy conservation in the respiratory chain by lanthanides provides a useful tool for studies on the mechanisms of $\Delta \overline{\mu}_{H^+}$ generation. It should be pointed out that the use of lanthanides is somewhat limited by their 'inconvenient' chemistry in aqueous solutions (limited solubility at neutral pH, strong tendency for complex formation, low solubility products with many anions) [11]. Our attempts to use lanthanides for coupling the ATP-dependent reactions have failed due to an immediate precipitation of their phosphate salts. Nevertheless, the results presented in the previous section show that, with some precautions, the lanthanides can be used for restoration of energy conservation in loosely coupled SMP.

The mechanism of the lanthanide-induced coupling

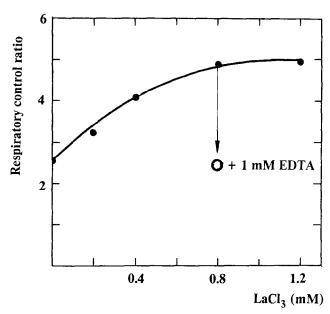


Fig. 2. An increase in the energy coupling capacity in submitochondrial particles in the presence of LaCl₃. The assay conditions are described in Fig. 1. The open point indicates the respiratory control ratio in the presence of 0.8 mM LaCl₃ and 1 mM EDTA.

remains to be established. Among several possibilities one seems to deserve brief discussion. The lanthanides are known to be powerful inhibitors of the mitochondrial Ca²⁺ uniport system [12,13]. It has been shown that the Ca²⁺ uniporter contributes significantly to the proton leakage in rat liver mitochondria, presumably due to the lack of absolute specificity of the Ruthenium red-sensitive cation transporting system [14,15]. The lanthanideinduced coupling may be caused by inhibition of the proton leakage through the Ca²⁺ uniporter. The absence of the Ruthenium red-induced coupling and much higher concentrations of lanthanides needed to increase the respiratory control (this paper), as compared to their efficiency in Ca²⁺ transport inhibition [16], may be explained by the opposite polarity of the coupling membrane in intact mitochondria and SMP. If the Ca²⁺ uniporter is asymmetrically arranged in mitochondria with the Ruthenium red-sensitive gate for Ca²⁺, H⁺ or lanthanides located on the outer surface of the inner membrane, the lack of the proton leakage sensitivity to Ruthenium red and low efficiency of lanthanides would be expected in inside-out SMP. The other mechanisms of the lanthanide-induced effect, such as phospholipid and/or protein immobilization, cannot be excluded. The latter explanation has been offered for the inhibitory effect of La⁺³ on

Table 1
Effect of La³⁺ and Dy³⁺ on the respiratory control ratio in submitochondrial particles

	NADH oxidase ^a (μmol/min per mg)		Respiratory control ratio
	-CCCP	+ 30 μM CCCP	-
Exp. 1			
1. Control ^b	0.25	0.86	3.5
2. +0.5 mM LaCl ₃	0.16	0.81	5.0
3. +0.5 mM LaCl ₃			
+20 mM succi- nate	0.29	0.96	3.4
Exp. 2			
1. Control ^b	0.38	1.06	2.8
2. +0.2 mM DyCl ₃	0.21	0.91	4.3
3. +0.2 mM DyCl ₃			
+0.5 mM EDTA	0.37	1.06	2.7
	Succinate oxidase ^c (µmol/min per mg)		
Exp. 3			
1. Control ^b	0.15	0.40	2.7
2. +1.2 mM LaCl ₃	0.07	0.35	5.0

^a Measured spectrophotometrically at 340 nm as described in Fig. 1. ^b Different batches of SMP treated with oligomycin (4 nmol/mg) to induce the respiratory control [2] were used.

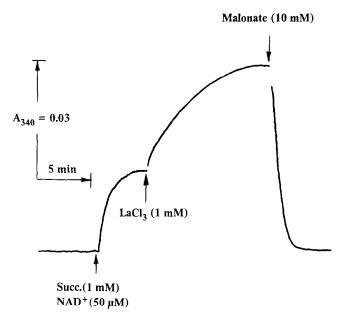


Fig. 3. Effect of LaCl₃ on the aerobic succinate-supported NAD⁺ reduction (reverse electron transfer). The reaction mixture (2 ml) contained 0.2 M sucrose, 50 mM Tris-acetate (pH 7.5), BSA (1 mg/ml) and submitochondrial particles treated with oligomycin (4 nmol/mg). SMP were pulsed with 5 μ M NADH before the reaction was initiated as indicated.

the slow phase of proton transfer during the bacteriorhodopsin photocycle [17].

Whatever the mechanism of the lanthanides' coupling effect, their capacity to increase energy conservation in the respiratory chain seems to be a useful experimental tool.

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^c Succinate oxidase was measured with an oxygen electrode in the mixture containing 0.2 M sucrose, 50 mM Tris-acetate (pH 7.5), BSA (1 mg/ml), 1 mM succinate and 5 μ M malonate (introduced with particles from the 'activation' medium [8]). The reaction was initiated by the addition of SMP (0.3 mg/ml).

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